

고속교반기와 주사기 펌프로 제조한 PCL과 PCL/PEG 마이크로캡슐의 약물방출거동

임현주 · 신지연 · 이득용[†] · 김배연* · 송요승**

대림대학교 의공융합과, *인천대학교 신소재공학과, **한국항공대학교 재료공학과
(2020년 2월 27일 접수, 2020년 4월 4일 수정, 2020년 4월 13일 채택)

Drug Delivery Behavior of PCL and PCL/PEG Microcapsules Prepared by High-speed Agitator and Syringe Pump

Hyunju Lim, Ji-Yeon Shin, Deuk Yong Lee[†], Bae-Yeon Kim*, and Yo-Seung Song**

Department of Biomedical Engineering, Daelim University, Anyang 13916, Korea

*Department of Materials Science and Engineering, Incheon National University, Incheon 22012, Korea

**Department of Materials Engineering, Korea Aerospace University, Goyang 10540, Korea

(Received February 27, 2020; Revised April 4, 2020; Accepted April 13, 2020)

초록: 1~5 μm 직경을 가진 니페디핀(NF)을 함유하는 폴리카프로락톤(PCL)과 PCL/폴리에틸렌 글리콜(PEG) 마이크로캡슐을 고속교반기를 이용하여 액중건조법으로 제조하여 교반속도, PCL 농도, PCL/PEG 중량비에 따른 캡슐의 형상과 약물전달거동을 조사하였다. PCL 농도가 8에서 2 wt%로 감소하고, 교반속도가 3000에서 5000 rpm로 증가함에 따라 캡슐크기는 감소하였다. 반면에, PCL/PEG 캡슐의 크기는 PCL/PEG 비가 10/0에서 6/4로 변화함에 따라 증가하였다. 직경이 작은 PCL 캡슐과 큰 PCL/PEG 캡슐에서 많은 양의 NF가 관찰되었다. FTIR 관찰결과, NF의 카보닐기와 아민기에 의해 PCL과 NF가 성공적으로 결합되어 있었다. 세포독성 실험결과, PCL과 PCL/PEG 캡슐은 세포 용해나 독성이 없어 안전하였다.

Abstract: Nifedipine (NF)-loaded PCL and PCL/PEG microcapsules with diameters ranging from 1 to 5 μm are synthesized by the oil-in-water emulsion-solvent evaporation technique to evaluate the influence of stirring speed, PCL concentration, and PCL/PEG weight ratio on morphology and drug delivery behavior of the capsules. The size of PCL capsules decreased as the PCL concentration decreased from 8 to 2 wt% and the stirring speed increased from 3000 to 5000 rpm. The diameter of PCL/PEG capsules increased by varying the PCL/PEG ratio from 10/0 to 6/4. Higher amount of released NF is observed for smaller PCL capsules and larger PCL/PEG capsules, respectively. FTIR results revealed that the NF-loaded PCL and PCL/PEG capsules were formed due to hydrogen bonding between PCL and NF. The capsules showed no evidence of cytotoxicity, suggesting that the capsules were safe.

Keywords: poly(ϵ -caprolactone) (PCL), poly(ethylene glycol) (PEG), nifedipine, microcapsule, drug release.

Introduction

Microcapsules consist of two parts: core and shell material. They have a structure in which solid or liquid or gas material (core) is encapsulated with a membranous material (shell). Core material containing an active ingredient can be extracted by breaking the shell to provide controlled release of drug with various means such as pressure, heat, light, acid, or chemical reaction.¹⁻⁷ The desired location can be reached at the ther-

apeutic level, making it a very useful carrier because the shell material shields the core material from the external environment and allows good release characteristics.²⁻⁷

Lung-targeted drug delivery systems (LTDDSs) are designed to deliver drugs to the disease site without causing adverse effects in other tissues and improve the therapeutic efficiency of the lung diseases such as asthma, tuberculosis, pneumonia, and lung cancer.⁸ Drug is likely to be delivered either via pulmonary inhalation or intravenous injection. The inhalation method is a non-invasive route that provides efficient lung targeting due to large surface area for drug absorption without the first-pass metabolism and the side effects. Particle size of drug or drug carriers play an important part in the particle stability

[†]To whom correspondence should be addressed.
duke1208@gmail.com, ORCID[®]0000-0003-1674-412X
©2020 The Polymer Society of Korea. All rights reserved.

in lung. Particles smaller than 1 μm and larger than 5 μm are mostly removed by exhalation and mucociliary, respectively. On the other hand, the particles with diameters ranging from 1 to 5 μm are found to be the optimal size due to easy deposition in alveolar regions and fast drug delivery.⁸ It usually takes 20 h to reach the plateau regime of nifedipine (NF) drug release for the PCL/PVP and PCL/PEG capsules in the size range of 150 to 250 μm .⁵ In the present study, a high-speed agitator and a syringe pump are employed to obtain a tailored particle size of 1 to 5 μm and fast drug delivery behavior. Bashir *et al.* also reported that increased concentration of polymer and larger capsule size are ascribed to delay in drug release, implying that the polymer/drug ratio is likely to be adjusted to achieve the controlled drug release rate.⁹

Poly(ϵ -caprolactone) (PCL) is widely used as a shell material for DDS due to its biodegradability, chief cost, elasticity, and excellent biocompatibility.^{5,10} PCL and poly(ethylene glycol) (PEG) have been used as shell materials in microcapsules with proper composition ratio. NF, as a core material, is a medication used for angina, hypertension, Raynaud's phenomenon, and premature labor.^{1,5-7} It has low cost as well as a short body half-life, causing low drug bioavailability as a result of fast release.⁶ The NF-loaded PCL and PCL/PEG microcapsules can be the practical approach to gain its therapeutic efficacy through achieving the sustained drug-release rate and improving the drug bioavailability.⁵⁻⁷ Uniform microcapsules with diameters in the range of 1 to 5 μm are synthesized by the oil-in-water (O/W) emulsion-solvent evaporation (ESE) technique using a high-speed agitator and a syringe pump. The water-insoluble oil phase is formed by mixing of PCL, PEG, NF and dichloromethane (DCM) and then dispersed in polyvinyl alcohol (PVA) emulsifier solution to obtain the microcapsule having the controlled drug delivery.^{2,5} In the present study, NF is

microencapsulated and then the capsule size, the NF release rate, and the cytotoxicity of the PCL and PCL/PEG microcapsules are then investigated.

Experimental

Materials. PCL ($M_n=80000$), PVA ($M_w=85000\sim 124000$), and NF ($\geq 98\%$) are purchased from Sigma-Aldrich. PEG ($M_w=2000$) is purchased from Yakuri Pure Chemicals (Japan). NF is 1,4-dihydro-2,6-dimethyl-4-(2-nitrophenyl)-3,5-pyridinedicarboxylic acid dimethyl ester, $C_{17}H_{18}N_2O_5$. DCM (99.5%, Samchun Pure Chemical, Korea) is purchased as a solvent and used as received without further purification.

Microcapsule Synthesis. The 2~8 wt% PCL solutions dissolved in DCM are prepared by stirring for 24 h at a speed of 500 rpm and then mixed with 10 mg NF to obtain a homogeneous solution. PCL and PCL/PEG solutions containing weight ratios in the range of 10/0 to 6/4 are prepared. PVA dissolved in distilled water is refluxed at 500 rpm and 80 $^{\circ}\text{C}$ to obtain a viscous homogeneous solution.⁵ The as-prepared PCL and PCL/PEG solutions are placed in a 20 mL B-D luer-lok syringe attached to the syringe pump (KDS-200, Stoelting Co., USA) and are fed into the 22-gage blunt-end metal needle at a flow rate of 0.25 mL/min.¹¹⁻¹⁴ They are injected into PVA aqueous solution (1.0 wt%) and stirred with a high-speed agitator (HD-312, HsiangTai, China) for 4 h at speed in the range of 3000 to 5000 rpm to obtain the desired microcapsules, as shown in Figure 1. The microcapsules are screened through a 25 μm sieve. The capsules are then cleaned in ethanol solution for 0.5 h and subsequently in distilled water for 0.5 h to remove the remnant solvent. The microcapsules are dried overnight in a vacuum oven. Prior to use, their storage and handling are conducted cautiously without light due to

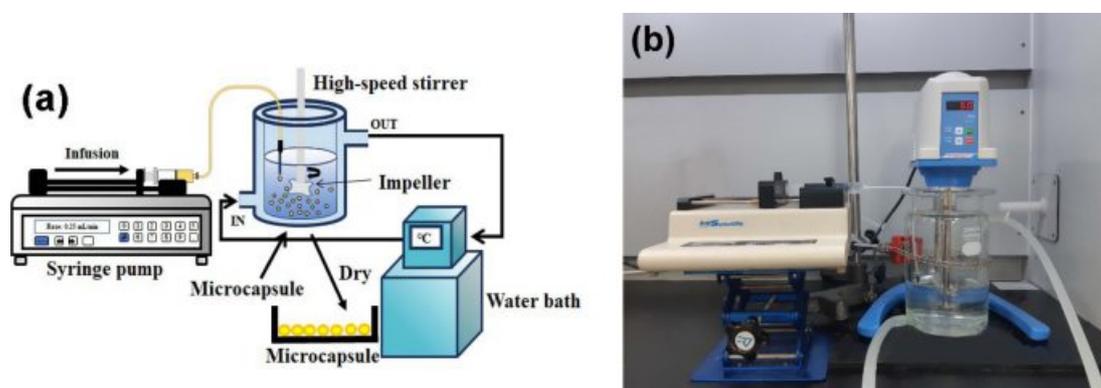


Figure 1. (a) A schematic diagram; (b) photograph of experimental apparatus.

photosensitivity of NF.⁵⁻⁷ The as-dried microcapsules are examined by using a SEM (S-3000H, Hitachi, Japan) and an optical microscope (SV-55, Sometech, Korea) equipped with *iSolution Lite image software* to investigate the microcapsule size and morphology. For the SEM observation, the microcapsules are sputtered with Au/Pd to ensure higher conductivity.¹¹⁻¹⁹

Characterization. NF (10 mg) and dried microcapsules (0.1 g) are dissolved in 20 mL ethanol by stirring in a 50 mL conical tube at 100 rpm and 37 °C. The samples (4 mL) are taken regularly throughout the experiment from the supernatant of the dissolved solution and then filtered through a 0.45 µm Millipore filter to remove the impurities. The adsorption at 238 nm is monitored to identify the variation in NF concentration using an UV-vis spectrophotometer (Jasco V-670, Japan). The amount of drug release is examined to evaluate the drug release rate as a function of time.⁵ The chemical properties of the capsules are analyzed by using the Fourier transform infrared spectroscopy (FTIR, Spectrum Two, PerkinElmer, UK). All experiments are carried out in triplicate. Values in the text are stated as the means±standard deviation, and $p < 0.05$ is weighed statistically significant.⁵

Cytotoxicity. The extract test method is conducted on the PCL and PCL/PEG microcapsules to evaluate cytotoxicity quantitatively according to the International Organization for Standardization (ISO 10993-5).^{5,13-19} The PCL and PCL/PEG microcapsules are extracted aseptically in single strength minimum essential medium with serum.⁵ The ratio of the microcapsules to the extraction vehicle is 0.2 g/mL (ISO 10993-12). The test extracts are placed onto three separate confluent monolayers of L-929 (NCTC Clone 929, ATCC, USA) mouse fibroblast cells propagated in 5% CO₂. EZ-cytox yields a water-soluble formazan, which can be provide different absorption spectra of the formed formazans. The absorbance of the colored solution is quantified by measuring at a wavelength of 415 nm with the microplate absorbance spectrophotometer (Bio-Rad, USA). Detailed experimental procedure is described elsewhere.^{5,13-19}

Results and Discussion

The effect of PVA emulsifier concentration on the size of the 4 wt% PCL microcapsule prepared by a magnetic stirrer is previously reported.⁵ For microcapsules prepared by using PVA emulsifier, uniform and highly stable spherical PCL, PCL/PVP, and PCL/PEG microcapsules (~150 µm) with a narrow

size distribution are successfully synthesized.⁵ This may be due to high affinity of the PVA for the shell materials and the core material.⁵ The NF release rate of PCL/PVP and PCL/PEG capsules increases with increasing PVP and PEG concentration due to the increase in capsule size. The 1 wt% PVA emulsifier is determined to be the best composition for the formation of uniform microcapsule. In the present study, PCL and PCL/PEG microcapsules using 1 wt% PVA emulsifier are studied by the ESE technique using the high-speed agitator. The PCL capsule size decreases gradually from 5.95±0.85 µm to 1.95±0.25 µm with increasing the stirring speed from 3000 to 5000 rpm. The microcapsule size is controlled by the balance between the turbulence to break the microcapsules, which are emulsion particles stabilized by PVA, and the internal viscosity and interfacial tension to hold the microcapsules. Fast stirring speed increases the turbulent energy acting on each microcapsule, increasing the rate at which the capsules becomes smaller, thereby reducing the size of the microcapsules.⁴ The capsules prepared at 5000 rpm are selected in the present study. In addition, the influence of PCL concentration on the capsule size is investigated. As the PCL concentration rises from 2 to 4, 6, and 8 wt% at a fixed speed of 5000 rpm, the PCL capsule size increases from 1.95±0.25 to 5.79±0.69, 7.08±0.98, 8.51±1.37 µm due to higher loading of polymer,⁵ as depicted in Figure 2. In the present study, the 2 wt% PCL capsule prepared at a stirring speed of 5000 rpm is chosen due to the capsule size (~1.95 µm) for LTDDS because carriers with diameters ranging from 1 to 5 µm are known to be responsible for easy deposition in alveolar regions.⁸

It is reported that melting temperature (T_m) of the PCL/PEG capsules rises from 65.0 to 65.5 °C with increasing the PEG content from 0 to 20 wt%.⁵ However, it decreases sharply down to ~62.4 °C when PEG is added to PCL more than 30 wt%. It is due to the blending effect of PEG because T_m of PEG is in the range of 50 to 52 °C. Although T_m of PCL/PEG capsules in the range of 60.0 to 62.4 °C is lower than those of the above-mentioned capsules (62.4–65.5 °C) having a diameter of about 154 µm, similar variation in T_m is observed as expected. T_m increases from 60.0 to 62.4 °C with increasing the PEG content from 0 to 20 wt% and then decreases down to 61.2 °C with further PEG doping, as shown in Figure 3. As the PCL/PEG weight ratio varies from 10/0 to 6/4, the capsule size increases gradually from 1.95±0.25 to 5.48±1.6 µm due to the hydrophilic PEG, as shown in Figure 4. The size of PCL/PEG capsules with a ratio of 10/0 to 8/2 is between 1 and 5 µm, which is a relevant capsule size for LTDDS.⁸ The affinity to

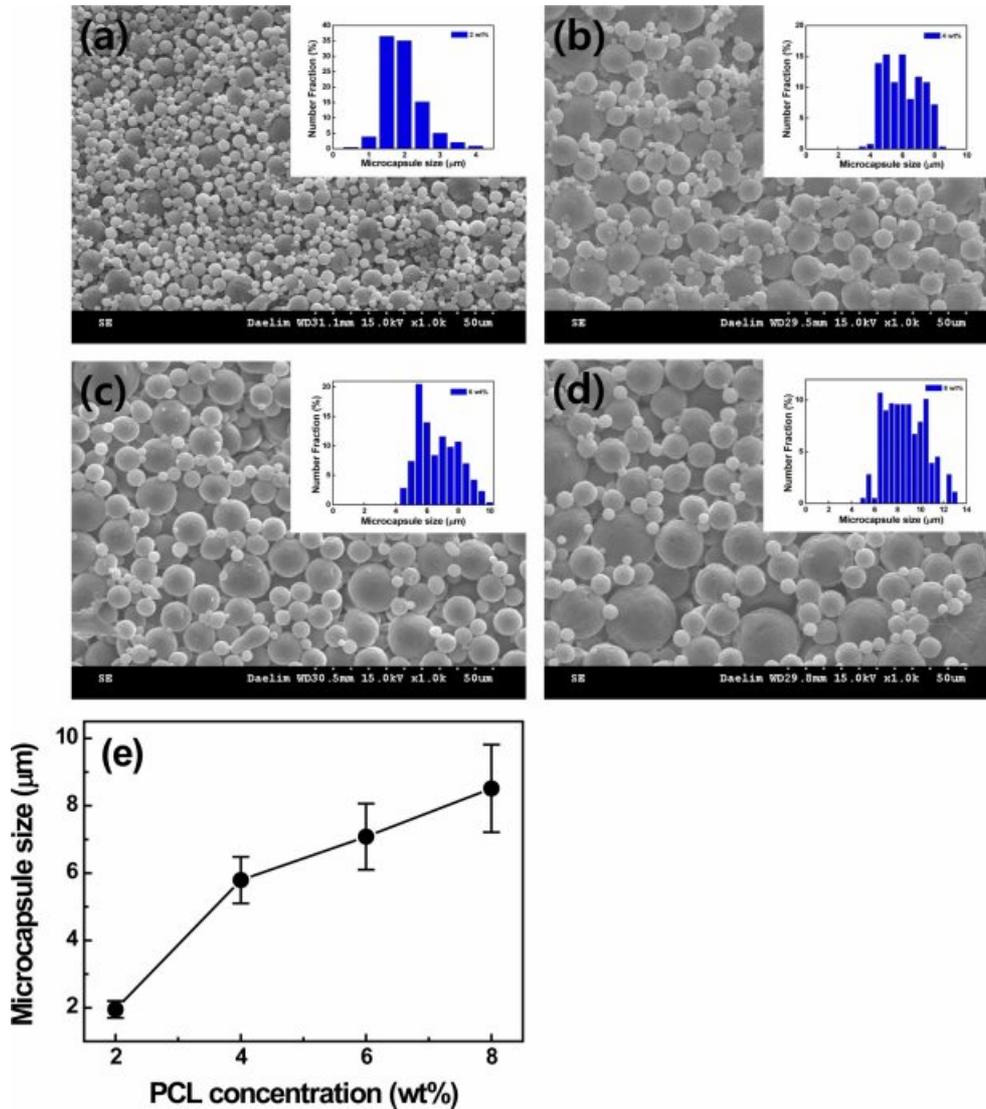


Figure 2. SEM images of (a) 2 wt%; (b) 4 wt%; (c) 6 wt%; (d) 8 wt% PCL microcapsules; (e) capsule size as a function of PCL concentration, respectively. Note that the microcapsules are prepared by using 1 wt% PVA emulsifier at a speed of 5000 rpm.

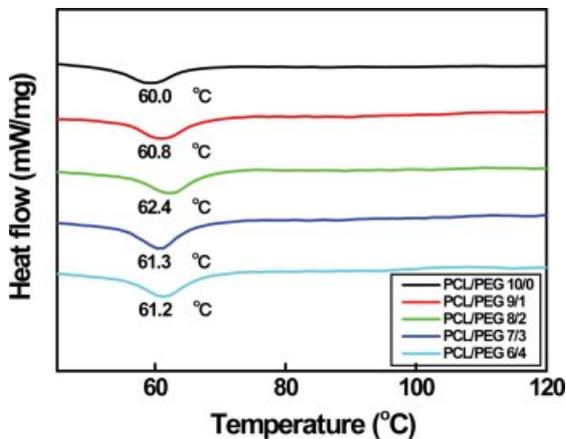


Figure 3. DSC curves of various PCL/PEG microcapsules.

water increases with increasing the content of the hydrophilic PEG, resulting in an increase in capsule size.⁵ As the PCL/PEG weight ratio varies from 10/0 to 6/4, the capsule morphology is changed from uniform spheres to agglomerated particles and airy bread-like distorted shape with a broad size distribution due to higher porosity.^{5,20} However, no relationship between capsule size and T_m of PCL/PEG is found.

The PCL spectrum exhibits the main peak at 1726 cm^{-1} corresponding to the carbonyl group ($-\text{CO}$) of the ester group. Peaks at 2937 and 2866 cm^{-1} corresponding to the asymmetrical and symmetrical methylene groups ($-\text{CH}_2$) are clearly visible.^{5,21} Peaks located at 1472 , 1412 , and 1361 cm^{-1} correspond to CH_2 bending. Bands at 1295 , 1230 , and 1164 cm^{-1}

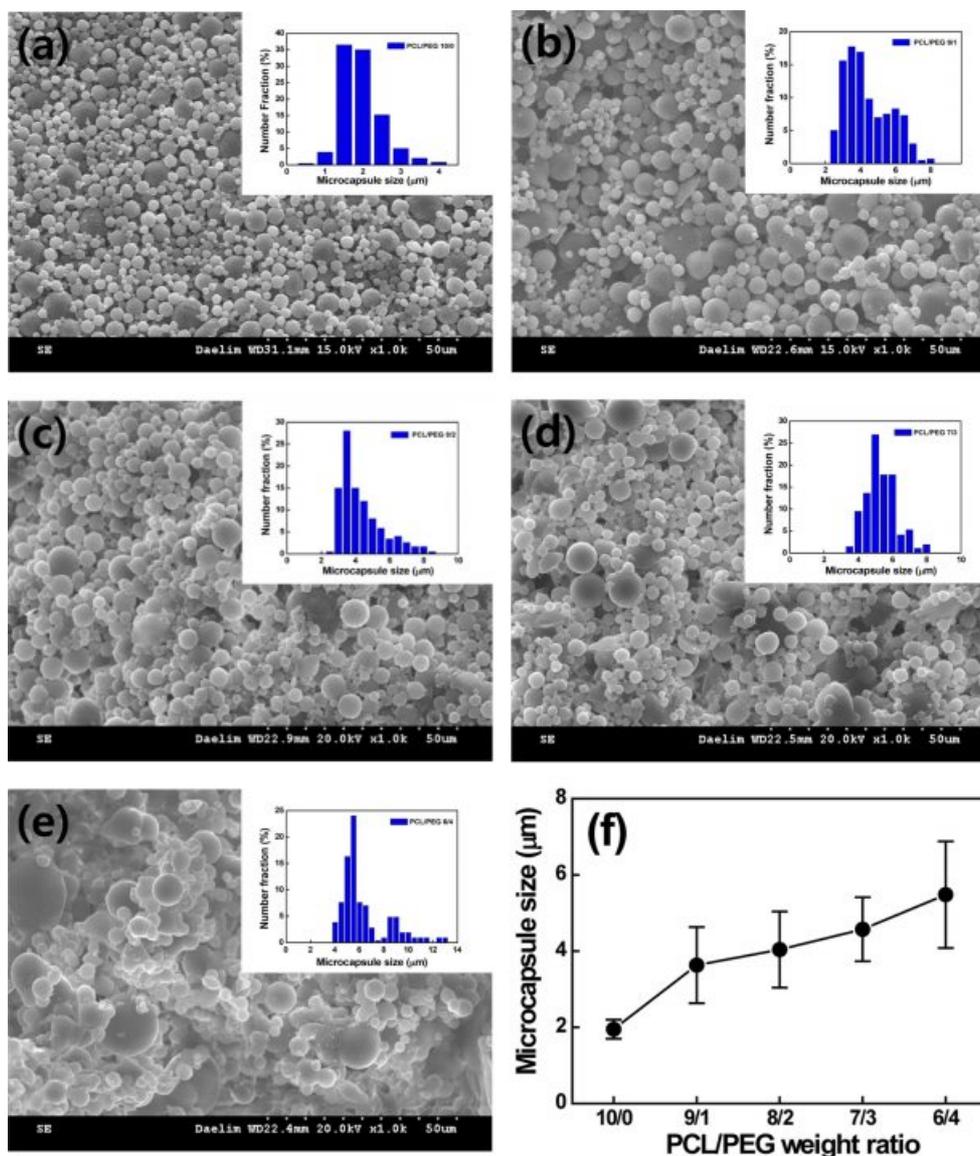


Figure 4. SEM images of microcapsules containing various PCL/PEG weight ratios of (a) 10/0; (b) 9/1; (c) 8/2; (d) 7/3; (e) 6/4; (f) capsule size as a function of PCL/PEG weight ratio, respectively.

related to C-C stretching, asymmetric and symmetric C-O-C stretching are observed, respectively.²¹ NF can potentially act as either proton acceptor (through the carbonyl groups, -CO) or proton donor (through the amine group, -NH).^{5-7,10} The hydrogen bonds (H-bonds) are reported to be formed via the amine group and the carbonyl groups. The H-bonded NF amine and carbonyl groups are observed at 3330 cm^{-1} (-NH) and at 1680 cm^{-1} (-CO), as depicted in Figure 5. The H-bonds formed among NF molecules are altered by those formed between NF and PCL. A sharp stretching vibration band at 1720 cm^{-1} is clearly observed on the NF-loaded PCL capsule

because of the NF carbonyl group formed H-bonds with PCL. A broad band situated in 3050 and 3700 cm^{-1} is visible in the spectra of NF-loaded PCL capsule, suggesting that the NF amine group associated with PCL via H-bond is established.⁵⁻⁷ The peak intensity of NF-loaded PCL/PEG capsule located at 1246 cm^{-1} (C-O-C) increases slightly compared to that of NF-loaded PCL capsule due to the presence of PEG, as shown in Figure 5. However, no appreciable change in FTIR peak is detected for the PCL/PEG (6/4) capsules. Typical PCL XRD peaks located at $2\theta = 21.4^\circ$, 22° , and 23.7° , corresponding to the (110), (111), and (200) planes of an orthorhombic crystal,

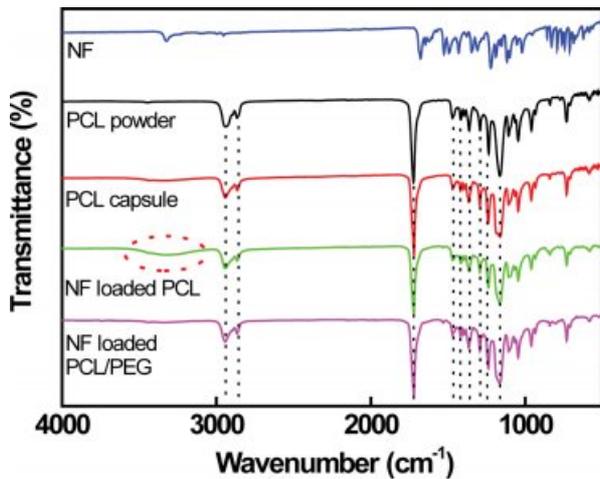


Figure 5. FTIR spectra of NF, PCL powder, PCL capsule, NF-loaded PCL, and PCL/PEG (6/4) capsules, respectively.

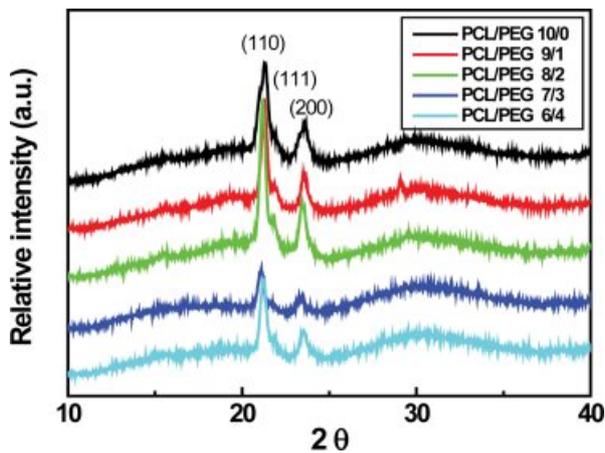


Figure 6. XRD patterns of PCL/PEG microcapsules having different concentration.

are clearly visible in Figure 6.^{5,23} These sharp peaks are attributed to the crystalline phase of PCL, which originates from the ordering of polymer side chains due to the intermolecular interaction between PCL chains through the hydrogen bonding.²³ Characteristic PEG peaks at 19.23° and 23.34° are reported.^{5,24} The (110) and (200) PCL peaks are shifted slightly to lower angle. In addition, the (110) PCL peak intensity starts to decrease with further addition of PCL/PEG ratio more than 7/3 probably due to the enhanced PEG contribution to the PCL/PEG microcapsules, as demonstrated earlier in Figure 5.⁵

The drug release behavior of NF is shown in Figure 7. Our previous studies reveal that NF release rate of PCL/PEG capsules prepared by a magnetic stirrer increases significantly

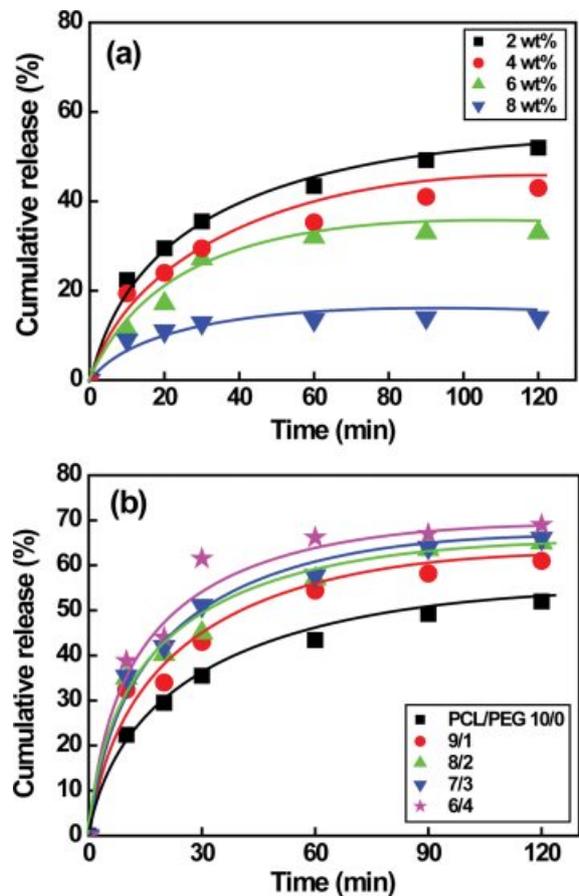


Figure 7. NF release behavior of (a) PCL; (b) PCL/PEG microcapsules. Note that the capsules are prepared by a high-speed agitator at a speed of 5000 rpm.

from 0 to 4 h at the beginning and then reaches the plateau region in 20 h probably due to larger capsule size in the range of 154 to $248 \mu\text{m}$.⁵ However, the drug release rate of PCL microcapsule made by a high-speed agitator and a syringe pump initially increases rapidly from 0 to 10 min in a short time and then reaches the plateau region in 1 h. The time reaches the plateau regime is 20 times faster than that of capsules prepared by a magnetic stirrer. The amount of released drug rises significantly from 14 to 52% with decreasing the PCL concentration from 8 to 2 wt% due to higher specific surface area as a result of the reduced capsule size from 8.51 ± 1.37 to $1.95 \pm 0.25 \mu\text{m}$, as shown in Figure 7(a). Unlike PCL capsules, Figure 7(b) suggests that the drug release rate of PCL/PEG capsules increases from 52 to 69% as the weight ratio of PCL/PEG varies from 10/0 to 6/4 (1.95 ± 0.25 to $5.48 \pm 1.6 \mu\text{m}$), implying that larger PCL/PEG capsules are attributed to higher amount of encapsulated drug.^{5,20} Although the capsule size

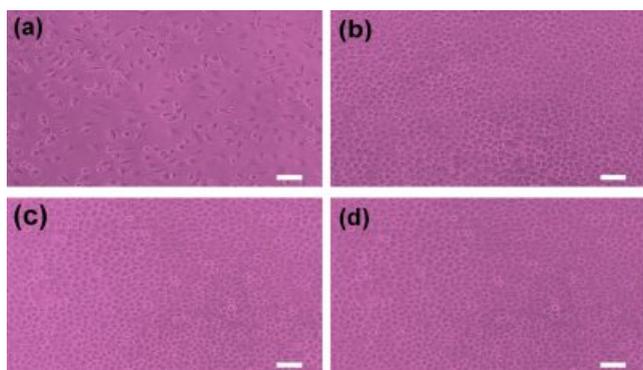


Figure 8. Photographs of cell morphologies: (a) positive control; (b) negative control; (c) PCL; (d) PCL/PEG microcapsule with NF, respectively. Scale bar is 50 μm .

affects the amount of drug, the drug release behavior remains constant regardless of the capsule size and the PEG concentration. The 2 wt% PCL and PCL/PEG (10/0 to 8/2) microcapsules with diameters ranging from 1 to 5 μm release the encapsulated NF from 52% to 65%.

A cytotoxicity test of PCL and PCL/PEG capsules determines whether a product or compound may have an adverse effect on tissues or cells.¹³⁻¹⁹ The test extracts with PCL and PCL/PEG capsules with NF show no evidence of causing cytotoxicity, as depicted in Figure 8. All experiments are performed in triplicate. The PCL and PCL/PEG (6/4) capsules exhibited quantitative cell viability of 121% and 133% compared to the negative control, respectively, as measured at a wavelength of 415 nm by using the iMark microplate absorbance spectrophotometer. The cell viability of PCL/PEG capsule is slightly higher than that of PCL capsule due to excellent biocompatibility of PEG.²⁰⁻²² The qualitative morphological grading of cytotoxicity of PCL and PCL/PEG capsules is determined to be scale 0.⁵ Therefore, it is conceivable that the NF-loaded PCL and PCL/PEG microcapsules have no cytotoxicity throughout the experiment and are considered to be clinically safe.

Conclusions

The NF-loaded PCL and PCL/PEG microcapsules are prepared by ESE technique through proper adjustment of the process parameters. The NF release rate of PCL and PCL/PEG microcapsules increases greatly from 0 to 10 min in a short time and then reaches the plateau region in 1 h. The amount of released drug rises drastically from 14 to 52% with decreasing the PCL concentration from 8 wt% ($8.51 \pm 1.37 \mu\text{m}$) to 2 wt%

($1.95 \pm 0.25 \mu\text{m}$) due to higher specific surface area. Unlike PCL capsules, the amount of encapsulated drug in PCL/PEG capsules increases with increasing the capsule size due to the increased hydrophilic PEG contribution to the PCL/PEG capsules. The PCL and PCL/PEG capsules exhibiting no evidence of cytotoxicity suggest that the microcapsules are clinically suitable for DDS.

Acknowledgment: This work was supported by the Materials & Components Technology Development Program (Project No. 20003560), funded by the Ministry of Trade, Industry & Energy (MOTIE, Korea).

References

1. M. Jang, C. Choi, W. Kim, Y. Jeong, and J. Nah, *Polym. Korea*, **28**, 291 (2004).
2. S. Kang, M. Baginska, S. R. White, and N. R. Sottos, *ACS Appl. Mater. Interfaces*, **7**, 10952 (2015).
3. N. V. N. Jyothi, P. M. Prasanna, S. N. Sakarkar, K. S. Prabha, P. S. Ramaiah, and G. Y. Srawan, *J. Microencapsul.*, **27**, 187 (2010).
4. Y. A. Kim, S. H. Kim, J. S. Park, D. S. Lee, J. G. Kim, and J. S. Shin, *J. Adhes. Interf.*, **13**, 17 (2012).
5. H. Lee, D. Y. Lee, Y. Song, and B. Kim, *J. Biomed. Eng. Res.*, **40**, 7 (2019).
6. J. Huang, R. J. Wigent, and J. B. Schwartz, *J. Pharm. Sci.*, **97**, 251 (2008).
7. C. J. Lee, H. J. Ha, S. Y. Kim, J. Y. Park, N. K. Jang, J. E. Song, and G. Khang, *Polym. Korea*, **39**, 739 (2015).
8. S. Nejati, E. M. Vadeghani, S. Khorshidi, and A. Karkhaneh, *Eur. Polym. J.*, **122**, 109353 (2020).
9. S. Bashir, M. Asad, S. Qamar, F. U. Hassnain, S. Karim, and I. Nazir, *Trop. J. Pharm. Res.*, **13**, 505 (2014).
10. J. Park, J. Kim, and N. Jeong, *Appl. Chem. Eng.*, **26**, 341 (2015).
11. H. Lee, D. Y. Lee, M. Lee, B. Kim, and Y. Song, *J. Electroceram.*, **42**, 124 (2019).
12. Y. Song, Y. Kim, D. Y. Lee, M. Lee, and B. Kim, *J. Nanosci. Nanotechnol.*, **17**, 7943 (2017).
13. B. Seol, J. Shin, G. Oh, D. Y. Lee, and M. Lee, *J. Biomed. Eng. Res.*, **38**, 248 (2017).
14. G. Oh, J. Rho, D. Y. Lee, M. Lee, and Y. Kim, *Macromol. Res.*, **26**, 48 (2018).
15. J. Kim, D. Y. Lee, E. Kim, J. Jang, and N. Cho, *Tissue Eng. Regen. Med.*, **11**, 32 (2014).
16. H. Jeong, J. Rho, J. Shin, D. Y. Lee, T. Hwang, and K. J. Kim, *Biomed. Eng. Lett.*, **8**, 267 (2018).
17. S. Son, J. Choi, H. Cho, D. Kang, D. Y. Lee, J. Kim, and J. Jang, *Polym. Korea*, **39**, 323 (2015).
18. J. Shin, H. Jeong, and D. Y. Lee, *J. Biomed. Eng. Res.*, **39**, 161 (2018).

19. S. Kim, J. Shin, Y. Yun, D. Y. Lee, D. Yang, B. Kim, and Y. Song, *Polym. Korea*, **43**, 764 (2019).
20. Y. Li, C. Zhu, D. Fan, R. Fu, P. Ma, Z. Duan, X. Li, H. Lei, and L. Chi, *Macromol. Biosci.*, **19**, e1800424 (2019).
21. E. Bolaina-Lorenzo, C. Martinez-Ramos, M. Monleon-Pradas, W. Herrera-Kao, J. V. Cauich-Rodriguez, and J. M. Cervantes-Uc, *Biomed. Mater.*, **12**, 015008 (2017).
22. N. S. V. Capanema, A. A. P. Mansur, A. C. de Jesus, S. M. Carvalho, L. C. de Oliveira, and H. S. Mansur, *Intl. J. Biol. Macromol.*, **106**, 1218 (2018).
23. M. Ravi, S. Song, J. Wang, X. Tang, and Z. Zhang, *Ionics*, **22**, 661 (2016).
24. C. Wang, L. Feng, H. Yang, G. Xin, W. Li, J. Zheng, W. Tian, and X. Li, *Phys. Chem. Chem. Phys.*, **14**, 13233 (2012).